

Observation on Quantitative and Qualitative Variability in Extracellular Enzymes of Certain Isolates of Genus *Alternaria*

Alka S. Petkar

Department of Botany, S. N. Arts, D. J. M. Commerce and B. N. S. Science College, Sangamner 422605, Maharashtra, INDIA.

Corresponding Address:

petkar_yoga@rediffmail.com

Research Article

Abstract: A few extracellular enzymes of the isolates of *Alternaria alternata* and *Alternaria triticina* were viscometrically determined. Quantitative variability was noticed in PG, PME, PGTE, PMTE, PMG, cellulase and protease in secretion in the culture medium by various isolates. Absence of PME activity in the culture filtrates of a few isolates of the same species was an interesting observation. Present findings point towards the fact that isolates of the same morphological species may differ in biochemical level.

Key words: Extracellular enzymes, *Alternaria alternata*, *Alternaria triticina*.

Introduction

The importance of vegetables in daily human diet is recognized all over the world. Vegetable are rich and comparatively cheaper source of vitamins and minerals. Consumption of these vegetables enhances taste, palatability, appetite and provides fibers for digestion. Vegetables play a key role in neutralizing all acids produced during digestion of fatty acids [1 & 3]. Even an urban householder observes in the course of his domestic chores that common vegetables are at times rotted and diseased. A good edible part of these vegetables is lost due to unsatisfactory conditions of transit and marketing [12, 29, 30 & 31]. Solanaceous fruits like so many other Fruits become soft and pulpy after attack of pathogens. Depression of tissue is common symptom of soft rot diseases. This depression in the tissue is due to collapse of cell wall skeleton. These symptoms are followed by maceration with the advancement of growth of pathogen. Maceration is due to degradation of middle lamella, which is made up of pectic substance [23]. Along with the pectic substance cellulose is also formed main skeleton of cell wall. In the degradation of pectic and cellulolytic substances pectic and cellulase enzyme play an importance role [4, 5, 8, 15, 11, 18, 20, 27 & 33]. Protein is also present in middle lamella a lay an important role in cell wall skeleton formation. The protein of middle lamella is hydrolysed by proteolytic enzymes [16 & 19]. These pectic, cellulolytic and protease enzymes degrade the skeleton of cell wall & convert complex molecules in

to simple molecules. These simple released molecules after hydrolysis of cell wall material is used as nourishment for pathogen. Thus extracellular enzymes play major key role in disease development by degradation of parenchymatous tissue of host [15, 26 & 32]. These extracellular enzymes are also secreted by pathogenic fungal organism in culture filtrates [16, 33, 36, & 39]. Aim of this study was to observed variability in secretion of extracellular enzymes of different isolates of morphologically same genus *Alternaria*.

Material and Methods

Diseased solanaceous fruits, showing symptoms like soft rot, water soaked area and sunken blackish brown lesions with concentric rings, were collected from vegetable market in separate polythene bags and were collected and brought to the laboratory for further study.

Isolation, Purification and Maintenance of Pathogen

Diseased fruits as such or in small pieces were surface sterilized by 1% mercuric chloride solution for about half to one minute [21]. Surface sterilized fruits were washed with distilled sterilized water. Surface sterilized pieces were inoculated on petriplates containing potato dextrose agar. Inoculated petriplates were incubated at 28°C to 30°C temperature. Isolated fungal pathogen was purified by repeated subculturing. Purified pathogens were taken in to stock cultures on agar slant. During all these operation Perfect aseptic conditions were maintained [17]. For pathogenicity tests isolated fungal organisms were inoculated on fresh and healthy solanaceous fruits and the reisolated [18, 19, 26 & 37].

Enzymes Assay

Enzymes extracts were obtained by growing pathogens in cole medium [13]. Culture filtrates were used for assay the activity of Polygalacturonase (PG), Pectin methyl esterase (PME), Polymethyl Galacturonase (PMG), Polygalacturonate transeliminase (PGTE), Pectin methyl transeliminase (PMTE), Protease and cellulase.

Viscometric study were made for the detection for PG, PMG, PGTE, PMTE, Protease and cellulase by using standard viscometric procedure [18,19,20 & 39].

Activity of PME was measured by continuous titration method [18]. Component of reaction mixture for enzyme assay are mentioned in Table 1.

Name of the enzyme to be detected	Components of reaction mixture in sequence				
	Substrate	Distilled water	Buffer	Enzyme Extract	Temperature of reaction
PG	3.5 ml. 1.2 % NaPP	1 ml.	MacIlvaine PH 4.6 1.5 ml.	1.5 ml.	$30 \pm 1^{\circ}\text{C}$
PGTE	3.5 ml. 1.2 % NaPP	1 ml.	MacIlvaine PH 8 1.5 ml.	1.5 ml.	$30 \pm 1^{\circ}\text{C}$
PMTE	3.5 ml. 1.2 % pectin	1 ml.	MacIlvaine PH 4.6 1.5 ml.	1.5 ml.	$30 \pm 1^{\circ}\text{C}$
PMG	3.5 ml. 1.2 % pectin	1 ml.	MacIlvaine PH 8 1.5 ml.	1.5 ml.	$30 \pm 1^{\circ}\text{C}$
Cellulase	3.5 ml. 0.5 % Carbory methyl cellulose	1 ml.	MacIlvaine PH 5.6 1.5 ml.	1.5 ml.	$30 \pm 1^{\circ}\text{C}$
Protease	3.5 ml. 1.2 % gelatin	1 ml.	Boric acid Borax PH 8.5 1.25 ml.	2.0 ml.	$40 \pm 1^{\circ}\text{C}$
PME	3.5 ml. 1.2 % pectin	1 ml.	PH 5.5	4.5 ml.	$30 \pm 1^{\circ}\text{C}$

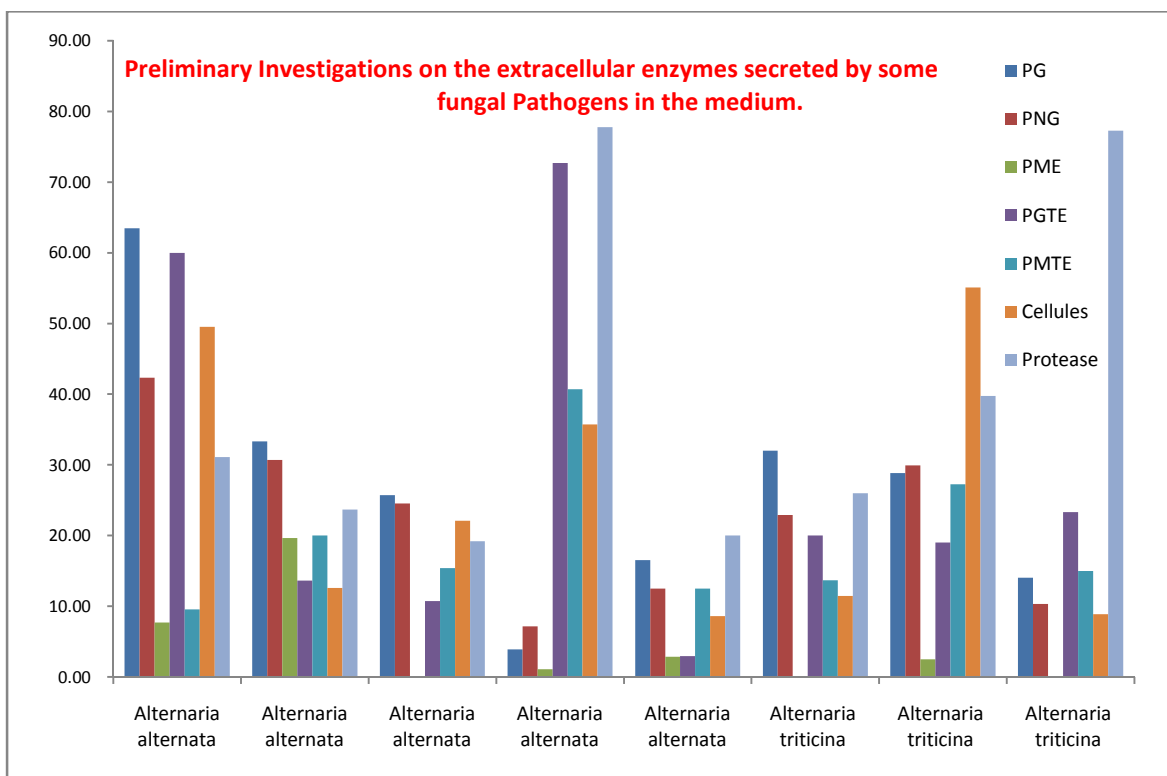
Result

All the isolates of genus *Alternaria* tested were able to produced extracellular enzymes in smaller or greater quantity except PME in few cases. Results are tabulated in Table-2.

- 1) **Polygalacturonase (PG)**- Secretion of this enzyme observed in all isolates of *Alternaria* Maximum amount was seen in *Alternaria alternata* Isolate No. 1 as compared to other isolates. Similarity Maximum amount of this enzyme was observed in *A. triticina* Isolate No. 6 as compared to other isolates of *A. triticina*.

Table 2: Preliminary Investigations on the extracellular enzymes secreted by some fungal Pathogens in the medium

S. No.	IMI No.	Iso-late	Name of Pathogen	Ph				Enzymes						
				Tm total	4 th day	7 th day	9 th day	PG	PNG	PME	PGTE	PMTE	Cellules	Protease
1.	203393	1	<i>Alternaria alternata</i>	5.2	4.5	7.7	7.8	63.48	42.31	7.70	60.00	9.57	49.53	31.11
2.	203394	2	<i>Alternaria alternata</i>	5.2	5.0	5.2	5.2	33.33	30.70	19.64	13.61	20.00	12.57	23.68
3.	203395	3	<i>Alternaria alternata</i>	5.2	5.0	5.2	5.2	25.70	24.54	--	10.71	15.38	22.11	19.18
4.	226407	24	<i>Alternaria alternata</i>	5.2	5.2	7.8	7.9	3.91	7.14	1.07	72.72	40.71	35.71	77.77
5.	203837	5	<i>Alternaria alternata</i>	5.2	5.1	5.2	5.2	16.51	12.50	2.86	2.94	12.50	8.59	20.00
6.	203838	6	<i>Alternaria triticina</i>	5.2	5.0	5.2	5.4	32.00	22.93	--	20.00	13.69	11.46	25.99
7.	203900	7	<i>Alternaria triticina</i>	5.2	5.0	5.2	6.2	28.82	29.91	2.50	19.03	27.27	55.09	39.77
8.	203808	8	<i>Alternaria triticina</i>	5.2	5.1	5.3	8.0	14.05	10.30	--	23.33	15.00	8.88	77.27



- 2) **Pectin Methyl esterase- PME** considerable amount of PME was secreted by *A. alternaria* Isolate No.2. Insignificant amount of this enzyme was found in *A. alternaria* Isolate No. 1, 2, 4 & 5 and *A. triticina* Isolate No.7, No. secretion of this enzyme was observed in *A. alternaria* Isolate No. 3 and *A. triticina* Isolate No. 6, 8 & 9.
- 3) **Polymethyl Transeliminase (PMTE)-** Maximum amount of PMTE was secreted by *Alternaria. alternata* Isolate No.24 as compared to other isolates of *A. alternata*. Similarly secretion of this enzyme was more in *A. triticina* isolate No.7 in comparison to other isolate of *A. triticina*.
- 4) **Polymethylgalacturonase (PMG)-** Maximum secretion of PME was observed in *A. alternata* isolate No.1 as compared to four other isolates of *A. alternata*. Considerable amount of secretion of this enzyme was observed in *A. triticina* isolate No.7 in comparison to other isolates.
- 5) **Polygalacturonase Trans- Eliminase PGTE** Maximum amount of PGTE was secreted by *A. alternata* isolate No. 24 as compared to other isolates of *A. alternata* & *A. triticina*.
- 6) **Cellulase-** It is significant to note that out of five isolates of *A. alternata*, isolate No. 1 was strongly cellulolytic similarly; secretion of cellulose was more in *A. triticina* isolate No. 7 as compared to other isolates of same species.

- 7) **Protease-** Maximum amount of protease was secreted by *A. alternata* isolate No. 4 as compared to other isolates. Similarly maximum secretion was observed in *A. triticina* isolate no. 8 as compared to other isolates of same species.

Discussion

Solanaceous fruits, like many other fruits become soft and pulpy after the attack of pathogenic fungi. The mechanism by which healthy parenchymatous tissue is converted in to soft and pulpy mass by pathogens remain almost completely unknown until later part of 19th century. In 1886 first time concluded that something moves in advance to the growth of fungal hyphae. [14]. Thus biochemical substances later on recognized as enzymes responsible for degradation of soft parenchymatous tissue of host [4, 8, 33, 39]. Many pathogenic and saprophytic organisms produce extracellular enzymes for degradation of cell wall material. Complete mechanism of working of enzymes on cell wall skeleton is well understood [14]. Pectic substances are hydrolyzed by pectic enzymes. Pectinmethylesterase liberates methyl group from pectin and convert it in to pectic acid which is hydrolyzed by polygalacturonase enzyme in to simple units. Cellulose is degraded by successive enzymatic action of cellulase secreted by microorganisms. The larger molecules of cellulose are hydrolysed in to simple and smaller units which are converted in to glucose [40]. Protein is

hydrolysed by protease enzyme in to simple structure [2, 22, 38]. Culture filtrates of pathogenic fungi tested in present study contain pectic, cellulolytic and proteolytic enzymes, which are capable of degrading cellulose, pectic, substance and proteolytic substances of cell wall of parenchymatous tissue of host converts healthy tissue. The action of these enzymes on parenchymatous tissue of host converts healthy tissue in to soft & pulpy mass. Association of extracellular enzymes with diseased tissue has also been confirmed [4, 5, 21, 34]. Role of these enzymes were also confirmed in pathogenesis [37]. It is significantly interesting to note that there is variation in secretion of extracellular enzymes quantitatively as well as qualitatively among the isolates of *A. alternata* and *A. tritici* of same morphological species. There are certain reports in favour of present conclusions. High degree of variability in production of extracellular enzymes of different isolates of same genus. *Puccinia graminis* (10). *Polyporus* Spp [37], *Fusarium oxysporum* [7], *Cochliobolus carbonum* [36], *Accremorium* Sp [24]. Variation in enzyme secretion by pathogenic fungal of same genus was also observed. [25,28]. Therefore quantitative & qualitative variations at Enzymative level among the isolates of same species suggest. The rethinking of status of species constructed only on the basis of morphological characters.

References

1. Abha Khetarpal and Kocchar G.K. (1991) Nutritional quality of fruits and vegetables and their importance in human health, new Renaissance Magazine.
2. Agriose G.N. plant pathology PP 57-80, Acedemic, Press, Newyork.
3. Akmal Khan and Tabassum Mamid (1986) Role of vegetables in human diet. Progressive farming, Vol.-6 No.4.
4. Batman, D.F. and Millar, R.L. (1966) Pectic enzymes in tissue degradation Ann., REv. Phytopathology 4, 119-146.
5. Batman D.F. (1966), Hydrolytic and trans eliminative degradation of pectic substances by extracellular enzymes of *Fusarium solani* F. phareli Phytopathology 56, 238-244.
6. Bill G.H. and J.D. Polishook, 1991 Microfungi *Carpinus caroliniana* can J. Bot.- 9, 1477-1482.
7. Bosland, W.P. and P.H. Willam 1987, An evaluation of *fusarium oxysporum* from Crucifers based on pathogenicity, iso enzyme polymorphism, vegetative compatibility and geography origin Can J. Bot-65, 2067-2073.
8. Brown, N 1965 Toxin and cell wall dissolving enzymes in relation to plant diseases. Ann rev. Phytopathology, 3, 1-18.
9. Burdon, J.J., N.H Luig and D.R. Marshall 1983. Isoenzyme uniformity and virulence variations in *Puccinia graminis* F.Sp. *tritici* and *P. recondita* F. Sp. *tritici* in Australia, Aust. J. Bot. 36, 403-410.
10. Burdon J.J. and A.P. Roelf 1985. The effect of sexual and Asexual reproduction on the isoenzyme structure of population *Puccinia graminis* Phytopathology 75, 1068-1073.
11. Cappellili R.A. (1966) Growth and Polygalacturonase production by *Rhizopus stolonifer*, Phytopathology 56, 734-737.
12. Chaudhary, B (1968), Diseases of vegetables during transit, storage and marketing and their control. Bull of the Indian Phytopathological Society No. 4, 65-70.
13. Cole, M. and Wood R.K.S. (1961) Pectic enzymes and phenolic substances in apples rotted by fungi Ann Botany 25, 435-452.
14. DeBary, A. (1986). Uber CInige Sclerotinlen and Sclerotinienkrankhelte Botan 2, 244; 377-474.
15. Demain, A. L. and Phaff, N. J. 1957 Recent advances in the enzymatic hydrlysis of pectic substances waleratein labs colluna 20, 119-140.
16. Ginsburge, B. Z. (1961) Evudances for a protein gel structure cross- linked by metal cations in the intercellular cement of plant tissue J. Expt. Bot. 12, 85-107.
17. Goel, S. K. and Mehrotra (1973) Production of pectolytic and cellulolytic enzymes by virulent and avirulent isolates of *Rhizoctonia Bataticola* (Trub) Butler in culture and in root extracts of *Abelmoschus* plants. Proc. Indian Nati. Sci. Acad. Part B. Biol Sci. 39 (6) 727-734.
18. Goodenongh, P. N. and Kempton, R. J. 1976. The activity of cell wall degrading enzymes in tomato root infected with *Pyrenochaeta Lycopersici* physiol. Plant Pathd 9 (13) 313-330.
19. Hancock, J.C. and Millar, R. L. 1965. Association of cellulolytic, proteolytic enzymes and xylolytic enzymes with Southern anthracnose, spring black stem, and *stemphylium* leaf spot of alfalfa. phytopathology, 55, 1061.
20. Hancock, J.H., Millar, R.L. and Lor Beer, J.W (1964) Pectolytic and cellulolytic enzymes produced by *Botrytis allii*, *B. cinerea* and *B. squamosa* in vitro and in vivo phytopathology 54, 923-931.
21. Hegde, S.V., A. Ramesha and C Srinvas (2011) Optimization of amylase production from and endophytic fungi Discosi sp isolated from *calophyllum inophyllum*, J. Agricultural Technol. 7 (3); 805-813.
22. Keen, N.T. (1966) Protease of *Pseudomonas lachryman* in relation to cucumber angular leaf spot phytopathology 56, 884 (Abstr.)
23. Kertesz, Z. I (1951), The pectic substances, Interscience Publishers new york. 628 P.
24. Leuchtmann, H. and A.K. CLay (1990) Isoenzyme Variations in the Acremonium / Epichloe Fungal Endophyte Complex Phytopathology 80; 1133-1139.
25. Levy, M.J. Romao, M.A. Marchetti and J.E. Hamer 1991. Fingerprinting with dispersed repeated sequences resolve pathotype diversity in the rice blast fungus plant cell 3; 95-102.
26. MacMillan, J.D. and Vaughn R.H. (1964) Purification and properties of a polygalacturonic Trans-eliminase produced by *Clostridium multifementan*, Biochemistry 3, 572-578.
27. Mehta, P., Vyas, K.M. and Sexena, S.B. (1976) Production of pectolytic enzyme by *Alternaria solani* and *Alternaria tenuis* on different culture media, J. Indian Bot Sci. 54; 200-206.
28. Michelmores, R.W. and S.H. Hulbert (1987) Molecular markers for genetic analysis of phytopathogenic fungi Ann. Rev. phytopathol 25; 383-404.
29. Ram A. and Lele V.C. (1968). A new leaf spot disease of brinjal caused by *Colletotrichum dematium* (Pers Ex. Fr.) Grover, Indian Phytopathology 127-130.
30. Ramsay, G.B. Naint, J.B. MacCollach, L.P. 1953 Market diseases of tomato, Peppers and Egg. Plants Handbook U.S. Dept. Agric No. 28 & 50.

31. Rangaswamy, G. and Sambandhan C.N. 1960, *Alternaria melongena* causing leaf spot & Fruit scab of egg. plant and fruit rot of chilli Mycologia 52; 517-520.
32. Rangaswamy, G. and Sambandhan C.N. 1961, Comparative studies on some *Alternaria* spp occurring on solanaceous hosts Indian J. Agric Sci. 31; 160-172.
33. Reese.E.T., Siu R.G.H. and Levinson H.S. (1960). The biological degradation of soluble cellulose derivatives and its relationship to the mechanism of cellulose hydrolysis J. Bacteriol-59 ; 435-497.
34. Sadasivam T.S. and Subramanian D. (1963) Pectic enzyme and plant diseases J. Indian Botany Soc. 42-A; 199-212.
35. Shannon M.C., S.K. Balial and J.W. Harris (1973), Starch gel electrophotoions J. Hered 76; 431-435.
36. Simcox K.D., D. Nickernt and W.L. Peterson (1992) Comparison of isoenzymes polymorphism in races of *cochliobolus carbonum* phytopathology-82; 621-624.
37. Singh G.P. and Husain A. (1968). Role of Enzymes in Pathogenesis by *Fusarium lateritium* *E. cajani* Indian phytopathology -31; 361-373.
38. Tookey P.W. and W.E. Fry and M.J. Villarreal Gonzales (1985) Isoenzymes characterization of sexual and Asexual *Phytophthora infestans* J. Hered – 76; 431-435
39. VanEtten H. and Batman D.F. (1965) Proteolytic activity of *Rhizoctonia* infected hypocotyls of bean phytopathology – 44; 1285 Abstr.
40. Whitakar D.R. (1957). The Mechanism of degradation of cellulose by *Myrothecium Verrucaria* cellulase, Can J. Biochem and physiol- 35; 733-742.
41. Wood R.K.S. (1960). Petic and Cellulolytic enzymes in plant diseases Ann. Rev. pl. Physiology -11; 299-322.